

## ANALYSIS

DETERMINATION OF LIPOPHILICITY AND  $pK_a$  MEASUREMENT OF SOME 4-IMINO-1,4-DIHYDROCINNOLINE-3-CARBOXYLIC ACID AND 4-OXO-1,4-DIHYDROCINNOLINE-3-CARBOXYLIC ACID DERIVATIVES – ISOSTERIC ANALOGUES OF QUINOLONESWIEŚŁAWA LEWGOWD<sup>a)</sup>, ANDRZEJ STAŃCZAK<sup>a)</sup>, ZBIGNIEW OCHOCKI<sup>a)</sup>  
and KRYSZYNA RZESZOWSKA-MODZELEWSKA<sup>b)</sup><sup>a)</sup> Department of Hospital Pharmacy, <sup>b)</sup> Department of Analytical Chemistry, Medical University of Łódź, Muszyńskiego 1, 90-151 Łódź, Poland

**Abstract:** As a continuation of our study, physicochemical properties of some antibacterial active 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives were investigated. Partition coefficient ( $\log P$ ), and dissociation constant ( $pK_a$ ) were experimentally determined and also calculated by the use of ACDLabs system software. The obtained values were correlated with experimental data.

**Keywords:** 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives, isosteric analogues of quinolones, partition coefficient ( $\log P$ ), RP-TLC,  $pK_a$  measurements.

The quinolone group of antibacterial agents is one of the most effective classes of drugs in the treatment of bacterial diseases. Cinoxacin, the chemical structure of which is characterized by a 4-oxocinnoline-3-carboxylic acid moiety shows also antibacterial activity, mainly against Gram-negative bacteria (1, 2). In our previous study, chemical modification of cinoxacin was studied with the aim of improving its antibacterial activity and spectrum (3). Therefore, the series of 4-imino-1,4-dihydrocinnoline-3-carboxylic acid derivatives were synthesized and their *in vitro* antibacterial activity against Gram-positive and Gram-negative bacteria were tested. The minimum inhibitory concentration (MIC) of the most active compounds was in the range of the first generation quinolones such as nalidixic acid. These derivatives were designed as isosteric analogues of fluoroquinolones and were characterized by the presence of an imine- group instead of an oxo-group at the 4-position and nitrogen atom in position 2. The chemical structure and minimum inhibitory concentration (MIC) of some of them are described in Table 1. Though it is known that quinolones are active against topoisomerases of type II and IV, the molecular details of their mode of action remain unclear (4). The variety of models propose binding of quinolones with DNA or non-covalent DNA-quinolone complexes (5). The chemical structure of synthesized compounds may suggest their antibac-

terial activity as potential intercalators. They have a planar polyaromatic system and ability to form hydrogen bond with DNA base. The tested compounds exhibit good polarizability as well and can be good electron acceptors to DNA base pairs.

Intercalators are the most important group of compounds that interact reversibly with the DNA double helix. The flat molecules of these ligands intercalate between pairs of DNA helix, lengthening and unwinding this structure at the intercalation sites. Intercalating agents share common structural features such as the presence of planar polyaromatic system, which binds by insertion between DNA base pairs, with a marked preference for 5'- pyrimidine – purine – 3' steps (6).

Six derivatives from the synthesized series were selected in our study to explain structure-activity relationships. The compounds were chosen considerably differing in the antibacterial activity to measure or calculate some physicochemical properties.

Biological activity of chemical compounds is associated with their physicochemical properties. The physicochemical constant (the dissociation constant  $K$  and partition coefficient  $\log P$ ) and some structural parameters (topological, steric and electronic) are useful for understanding the behavior of drug molecules. Therefore, the physicochemical parameters ( $\log P$  and  $pK_a$ ) of synthesized 4-imino-

1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives, influencing the biological availability were determined experimentally and calculated by the use of ACDLabs system software (7).

To evaluate the validity of  $pK_a$  constants predicted by the ACDLabs method, UV-visible spectroscopic measurements were made at different pH values (8). Spectra of **I-VI** were recorded in the range of 200–400 nm, as a function of  $A = f(\text{pH})$  and the  $pK_a$  value was calculated according to the Henderson-Hasselbalch equation. The obtained results are presented in Table 2.

The experimental  $pK_a$  values were correlated with those calculated (ACDLabs system software). The comparison of experimental and calculated data gave the equation:

$$\text{clog } pK_a = a pK_a + b \quad (1)$$

where:

$$a = 1.0281, b = -0.0757, r = 0.9977, s = 0.2387, F = 1279.013$$

Comparison of these results permits to define usefulness of computer software for calculation of physicochemical parameters of the compounds.

The partition coefficient ( $\log P$ ) is also a very functional parameter, which may be used in combination with  $pK_a$  to predict the distribution of tested compounds in biological systems as a measure of their lipophilicity. The relative lipophilicity of 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives **I-VI** were determined by reversed-phase thin-layer chromatography (RP-TLC). The chromatographic parameter of hydrophobicity in this method is the  $R_M$  value, defined as:

$$R_M = \log \left( \frac{1}{R_f} - 1 \right) \quad (2)$$

Its value decreases linearly with the increase of concentration of organic modifier (DMF) in the mobile phase. Correlation of both sets of values and extrapolation to zero values of the organic modifier (DMF) permits to obtain normalized chromatographic parameter  $R_{MW}$ .

The experimental lipophilicity values ( $\log P_{TLC}$ ) at pH 7.4 of all compounds **I-VI** were determined by the use of the calibration curve, which was obtained under the same measurement conditions for the set of six standards of known experimental lipophilicity (9). As the standards were used: acetanilide (**1**), 4-ethoxyacetanilide (**2**), 3,4-dichloraniline (**3**), benzophenone (**4**), 1-bromonaphthalene (**5**), and 1,2,3,4-tetrabromobenzene (**6**). Correlation of both sets of values gave the calibration equation:

$$\log P_{TLC} = 1.0428 R_{MW} - 0.1005 \quad (3)$$

$$r = 0.9997, F = 9662.0671, s = 0.03, n = 6$$

The literature values of  $\log P$  and  $R_{MW}$  of the standards are presented in Table 3.

Terms of the linear equations correlating the  $R_{MW}$  values of derivatives **I-VI** with the concentration of DMF [c (% , v/v)] in the mobile phase and calculated results of  $\log P_{TLC}$  (pH = 7.4) are presented in Table 4.

Because the distribution coefficient at the given pH is denoted  $\log D$ , the  $\log P_{TLC}$  value determined in this study, at pH 7.4, was denoted  $\log D_{7.4}$ . The experimental values of partition coefficient ( $\log P_{TLC}$ ) of compounds **I-VI** were correlated with the calculated values ( $\log P_{ACDLabs}$ ), which predicts the logarithm of the partition coefficient of compounds in *n*-octanol – water system, to define usefulness of computer software for calculation of the  $\log P$  parameters. The obtained values, compared with the experimental values of  $\log P_{TLC}$  ( $\log D_{7.4}$ ) are presented in Table 5. The comparison of experimental and calculated data gave the equation:

$$\text{clog } P = a \log P_{TLC} + b \quad (4)$$

where for ACDLabs system software:

$$a = 1.3277, b = -1.0986, r = 0.9600, s = 0.3737, F = 95.9456$$

## EXPERIMENTAL

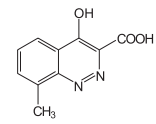
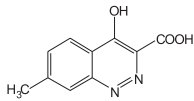
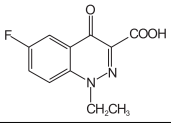
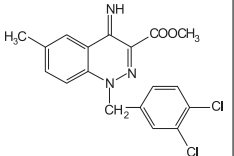
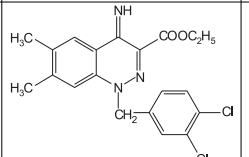
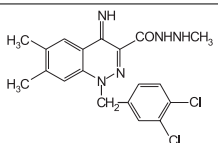
Chemistry and biological activity of tested compounds were described earlier (3).

UV spectrophotometric method of determination of  $pK_a$  values

The spectrophotometric measurements were recorded with double-beam spectrophotometer UV/VIS PAY-4 (Unicam, Denmark) with a fixed slit of 2 nm. A numerical pH-meter PHM-8 (Radiometer, Denmark) and a special glass electrode (type GK 2725 Radiometer, Denmark) for pH measurements in the  $1 < \text{pH} < 10$  range were used. The glass electrode was calibrated according to standard procedures with a phosphate buffer pH = 6.88, borate buffer pH = 9.22 and citrate buffer pH = 3.0 (Merck). The temperature was kept constant at 20°C. The UV spectra of all compounds were recorded in 1 cm quartz cells over the range 200 – 400 nm. The stability of the compounds was studied spectrophotometrically at concentration  $c \approx 1 \times 10^{-4}$  M in 1%, v/v aqueous solutions of DMF,  $\mu = 0.1$ .

Spectrophotometric method was selected due to its applicability in highly diluted solutions of compounds poorly soluble in water. Standard solutions were prepared by dissolving weighted amounts of

Table 1. Structure and MIC some of 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives.

| No. | Compound  | <i>In vitro</i> antibacterial activity of the compounds<br>[Minimum inhibitory concentration (MIC) µg/mL] |                                  |                                  |                                    |                                 |
|-----|---|---|----------------------------------|----------------------------------|------------------------------------|---------------------------------|
|     |   | <i>S. aureus</i><br>ATCC 6538 P   | <i>S. aureus</i> *<br>ATCC 29213 | <i>S. aureus</i> *<br>ATCC 25923 | <i>E. faecalis</i> *<br>ATCC 29212 | <i>B. subtilis</i><br>ATCC 6633 |
| I   |    | 25  | -                                | -                                | 25                                 | -                               |
| II  |    | -   | 25                               | 25                               | 25                                 | -                               |
| III |    | -   | -                                | -                                | -                                  | >400                            |
| IV  |   | 125   | 125                              | 125                              | 250                                | 125                             |
| V   |  | 25  | 25                               | 25                               | 50                                 | 25                              |
| VI  |  | 50  | 50                               | 50                               | 50                                 | 50                              |

\* multiple resistant bacteria

particular compounds (3 – 8 mg) in 1 mL of DMF, and then filling with water to 100 mL in calibrated flasks. The analyzed solutions were made from standard solutions by dilution. Then, spectra of A = f (pH) within the range 200-400 nm were recorded. In order to choose the optimal wavelength at which the difference in absorbance between the two forms is the biggest, absorption curves for dissociated and undissociated forms were drawn. The change in pH of the solutions was obtained using various concentrations of HClO<sub>4</sub> (c = 1.0; 0.1; 0.01; 0.001 M) and solutions of anhydrous NaOH (c = 1.0; 0.1; 0.01; 0.001 M).

#### Chromatographic system and conditions

Chromatography was carried out on TLC glass plates (5 cm × 10 cm) coated with RP-18 F<sub>254s</sub> (Merck,

Darmstadt, Germany). The mobile phase was DMF – aqueous buffer TRIS, of pH = 7.4 and ionic strength 0.2 M. The organic modifier (DMF) content varied from 40 to 85% (v/v) in 5% increments. Migration of 8.5 cm was obtained on all the plates by cutting the layer at 9.5 cm and spotting the compounds on a 1 cm line of the plate. Compounds were dissolved in methanol (1.5 mg/mL) and 2 µL of the solution was spotted on the plates. After developing and drying, the spots were observed under UV light and the R<sub>F</sub> values are mean from three determinations.

The experimental lipophilicity (log P<sub>TLC</sub>) of all compounds was determined by the use of the calibration curve (eq. 3), which was obtained under the same measurement conditions for the set of six standards of known experimental lipophilicity.

Table 2. Spectrophotometric determination of p*K*<sub>a</sub> values of 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives.

| Compd. No.              | Concentration [M]       | λ [nm] | p <i>H</i> | A    | p <i>K</i> <sub>a</sub> | p <i>K</i> <sub>a</sub> (average values) |
|-------------------------|-------------------------|--------|------------|------|-------------------------|--|
| 1                       | 2                       | 3      | 4          | 5    | 6                       | 7  |
| <b>I</b>                | 5.39 × 10 <sup>-6</sup> | 290    | 4.58       | 0.37 | 3.95                    | 4.08 ± 0.35                              |
|                         |                         |        | 4.00       | 0.41 | 3.72                    |  |
|                         |                         |        | 3.25       | 0.55 | 4.13                    |  |
|                         |                         |        | 3.09       | 0.57 | 4.49                    |  |
|                         |                         |        | 2.87       | 0.70 | 4.15                    |  |
|                         | 1.13 × 10 <sup>-4</sup> | 343    | 8.64       | 0.33 | 9.53                    | 10.10 ± 0.60                             |
|                         |                         |        | 9.42       | 0.39 | 10.02                   |  |
|                         |                         |        | 10.14      | 0.49 | 10.42                   |  |
|                         |                         |        | 10.60      | 0.67 | 10.10                   |  |
|                         |                         |        | 11.00      | 0.80 | 10.45                   |  |
| <b>II</b>               | 8.33 × 10 <sup>-4</sup> | 270    | 3.16       | 0.70 | 3.33                    | 3.50 ± 0.20                              |
|                         |                         |        | 3.69       | 0.75 | 3.42                    |  |
|                         |                         |        | 4.37       | 0.77 | 3.41                    |  |
|                         |                         |        | 4.60       | 0.80 | 3.70                    |  |
|                         |                         |        | 4.90       | 0.81 | 3.65                    |  |
|                         | 1.66 × 10 <sup>-4</sup> | 360    | 5.79       | 0.70 | 10.35                   | 10.38 ± 0.40                             |
|                         |                         |        | 6.44       | 0.68 | 10.05                   |  |
|                         |                         |        | 6.93       | 0.60 | 10.50                   |  |
|                         |                         |        | 7.57       | 0.50 | 10.29                   |  |
|                         |                         |        | 8.06       | 0.45 | 10.09                   |  |
| 5.42 × 10 <sup>-4</sup> | 275                     | 3.24   | 1.74       | 3.89 | 3.84 ± 0.20             |  |
|                         |                         | 3.54   | 1.56       | 3.71 |                         |  |
|                         |                         | 3.71   | 1.40       | 4.06 |                         |  |
|                         |                         | 3.90   | 1.24       | 3.85 |                         |  |
|                         |                         | 4.30   | 0.95       | 3.70 |                         |  |
| <b>IV</b>               | 5.20 × 10 <sup>-4</sup> | 365    | 4.23       | 0.38 | 3.60                    | 3.50 ± 0.30                              |
|                         |                         |        | 3.84       | 0.40 | 3.38                    |  |
|                         |                         |        | 3.13       | 0.47 | 3.34                    |  |
|                         |                         |        | 2.48       | 0.54 | 3.42                    |  |
|                         |                         |        | 1.70       | 0.55 | 3.42                    |  |
| <b>V</b>                | 5.20 × 10 <sup>-5</sup> | 362    | 4.10       | 0.80 | 4.24                    | 4.24 ± 0.08                              |
|                         |                         |        | 4.30       | 0.73 | 4.25                    |  |
|                         |                         |        | 4.57       | 0.65 | 4.32                    |  |
|                         |                         |        | 4.70       | 0.58 | 4.24                    |  |
|                         |                         |        | 4.96       | 0.50 | 4.18                    |  |
| <b>VI</b>               | 5.90 × 10 <sup>-5</sup> | 359    | 4.70       | 0.58 | 4.09                    | 4.01 ± 0.10                              |
|                         |                         |        | 4.41       | 0.55 | 4.00                    |  |
|                         |                         |        | 4.12       | 0.51 | 3.92                    |  |
|                         |                         |        | 3.90       | 0.45 | 4.02                    |  |
|                         |                         |        | 3.52       | 0.36 | 4.05                    |  |
|                         | 2.45 × 10 <sup>-5</sup> | 368    | 3.10       | 0.34 | 3.99                    | 9.01 ± 0.20                              |
|                         |                         |        | 9.95       | 0.58 | 9.18                    |  |
|                         |                         |        | 9.52       | 0.65 | 8.95                    |  |
|                         |                         |        | 9.25       | 0.72 | 9.07                    |  |
|                         |                         |        | 8.85       | 0.80 | 8.77                    |  |
|                         |                         |        | 8.40       | 0.95 | 9.05                    |  |

Table 3. Comparison of literature (log P) and experimental values of lipophilicity (R<sub>MW</sub>) for the standards used\*.

|                      | 1      | 2      | 3      | 4      | 5      | 6      |
|----------------------|--------|--------|--------|--------|--------|--------|
| log P                | 1.16   | 1.58   | 2.69   | 3.18   | 4.06   | 5.13   |
| R <sub>MW</sub>      | 1.19   | 1.62   | 2.71   | 3.10   | 3.98   | 5.02   |
| -b                   | 0.0214 | 0.0257 | 0.0354 | 0.0415 | 0.0512 | 0.057  |
| r                    | 0.9991 | 0.9977 | 0.9962 | 0.9978 | 0.9965 | 0.9949 |
| log P <sub>TLC</sub> | 1.14   | 1.58   | 2.72   | 3.13   | 4.05   | 5.13   |

\*acetanilide (1), 4-ethoxyacetanilide (2), 3,4-dichloraniline (3), benzophenone (4), 1-bromonaphthalene (5), and 1,2,3,4-tetrabromobenzene (6).

Table 4. Parameters of linear correlations between the R<sub>MW</sub> values of 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives and the concentration of DMF [c (%), v/v] in the mobile phase.

| Compd. No. | R <sub>MW</sub> | -b     | n  | r      | log P <sub>TLC</sub> (pH = 7.4) |
|------------|-----------------|--------|----|--------|---------------------------------|
| I          | 1.821           | 0.0235 | 9  | 0.9962 | 1.798                           |
| II         | 1.931           | 0.0321 | 8  | 0.9978 | 1.913                           |
| III        | 1.442           | 0.0231 | 8  | 0.9979 | 1.403                           |
| IV         | 3.460           | 0.0463 | 10 | 0.9957 | 3.507                           |
| V          | 4.502           | 0.0618 | 9  | 0.9964 | 4.594                           |
| VI         | 3.169           | 0.0427 | 9  | 0.9963 | 3.204                           |

Table 5. Comparison of the experimental and predicted values of physicochemical constants of 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives.

| Compd. No. | pK <sub>a</sub> (experimental) | pK <sub>a</sub> (ACDLabs)   | log P <sub>TLC</sub> (log D <sub>7.4</sub> ) | Log P (ACDLabs) |
|------------|--------------------------------|-----------------------------|--|-----------------|
| I          | 4.08 ± 0.35<br>10.10 ± 0.60    | 4.02 ± 0.50<br>10.40 ± 0.50 | 1.798  | 1.23 ± 1.02     |
| II         | 3.50 ± 0.20<br>10.38 ± 0.40    | 3.93 ± 0.50<br>10.49 ± 0.50 | 1.913  | 1.23 ± 1.02     |
| III        | 3.84 ± 0.20                    | 3.86 ± 0.20                 | 1.403  | 0.75 ± 0.69     |
| IV         | 3.50 ± 0.30                    | 3.53 ± 0.40                 | 3.507  | 4.14 ± 0.75     |
| V          | 4.24 ± 0.08                    | 3.91 ± 0.40                 | 4.594  | 4.60 ± 0.75     |
| VI         | 4.01 ± 0.10<br>9.01 ± 0.20     | 3.99 ± 0.40<br>9.27 ± 0.20  | 3.204  | 3.26 ± 0.75     |

## RESULTS AND DISCUSSION

### Chemistry

#### SAR analysis

For 4-imino-1,4-dihydrocinnoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinnoline-3-carboxylic acid derivatives **I-VI**, a comparative computer analysis was done. All the structures of the studied compounds were geometrically optimized by the use of semiempirical method INDO [algorithm Polak-Ribiere, RMS grad = 0.01 kcal/(Å mol), *in vacuo*, HyperChem 5.1] (10). The systematic conformational analysis was not used. The physicochemical

parameters of the compounds, received from ChemPlus modules of HyperChem are summarized in Table 6.

Geometric optimization of the studied compounds revealed a flat character of the basic core, what is the primary condition of intercalators structure. It seems that their biological activity is connected with:

1. A size of molecules, because many antibacterial compounds possess Van der Waals surface of molecule (GRID) above 300 Å<sup>2</sup> and similar volume, above 300 Å<sup>3</sup>. Thus their capacities are like other small organic molecules, applied as intercalating agents, such as naphthalimides.

Table 6. Some physicochemical parameters of 4-imino-1,4-dihydrocinoline-3-carboxylic acid and 4-oxo-1,4-dihydrocinoline-3-carboxylic acid derivatives.

| Compd. No. | Van der Waals area of molecule (GRID) [Å <sup>2</sup> ] | Van der Waals volume [Å <sup>3</sup> ] | Refractivity | Polarizability [Å <sup>3</sup> ] | Hydrogen bond acceptors* | Hydrogen bond donors* | Log P (ChemPlus) | Dipole moment [D] |
|------------|---|--|--------------|----------------------------------|--------------------------|-----------------------|------------------|-------------------|
| <b>I</b>   | 199.40  | 169.18                                 | 52.96        | 21.32                            | 5                        | 2                     | 1.96             | 2.61              |
| <b>II</b>  | 200.39  | 169.08                                 | 52.96        | 21.32                            | 5                        | 2                     | 1.96             | 3.10              |
| <b>III</b> | 227.22  | 190.20                                 | 57.44        | 21.84                            | 5                        | 1                     | 2.16             | 5.62              |
| <b>IV</b>  | 376.78  | 328.91                                 | 103.31       | 39.96                            | 6                        | 1                     | 5.50             | 3.55              |
| <b>V</b>   | 365.91  | 332.94                                 | 108.35       | 41.80                            | 6                        | 1                     | 5.97             | 6.79              |
| <b>VI</b>  | 377.34  | 337.55                                 | 108.85       | 42.03                            | 6                        | 3                     | 4.93             | 6.76              |

\*Interactive Analysis Predictor (<http://www.logp.com/>)

2. Higher values of refractivity and polarizability, which can determine the flexibility of these derivatives during interaction with biological target. This is connected with growing values of dipole moments as well.

3. Ability to create hydrogen bonds; more active compounds are good hydrogen bond acceptors.

4. Higher values of lipophilicity. Hydrophobic/hydrophilic properties of the compounds guarantee their effective transport and consequently the biological activity. The lipophilicity parameters of the synthesized compounds, determined by RP-TLC and expressed as log  $P_{TLC}$  (log  $D_{7,4}$ ) values are within the range 1.798-4.594 and effect the bioactivity of the derivatives. And though organic solvents with a log P between 1.5 and 4.0 are extremely toxic for microorganisms and other living cells, the most active compound **V** possesses higher log  $P_{TLC}$  (log  $D_{7,4}$ ) value.

A size of substituent at position 1. A nontypical dichlorobenzyl substituent with aromatic ring can be stacking with DNA bases, deciding about size and increasing lipophilicity of the molecule.

$pK_a$  and log P values of compounds **I-VI** were calculated as well by the use of the ACDLabs system software. The obtained results show a very good linear correlation ( $r > 0.95$ ). It can suggest that methodologies are suitable for dissociation constant and lipophilicity calculation.

This study was supported by the Medical University in Łódź, Poland, Research Programme No. 503 3011 3 and 503 3016 3.

## REFERENCES

1. The Merck Index, 12th edition, (Ed.: Susan Budavari), Merck Research Laboratories, Division of Merck & Co., Inc, Whitehouse Station, New York 1996.
2. Negwer M.: Organic-Chemical Drugs and their Synonyms (An International Survey) 7th ed: Akademie-Verlag GmbH, Berlin 1994.
3. Stańczak A., Ochocki Z., Martynowski D., Główna M., Nawrot E.: Arch. Pharm. Pharm. Med. Chem. 336, 18 (2003).
4. Drlica K., Zhao, X.: Microbiol Mol. Biol. Rev. 61, 377 (1997).
5. Siegemund K., Maheshwary S., Narayanan S., Connors W., Riedrich M., Printz M., Richert C.: Nucleic Acids Res. 33, 4838 (2005).
6. Braña M. F., Cacho M., Gradillas A., De Pasqual-Teresa B., Ramos A.: Curr. Pharm. Des. 7, 1745 (2001).
7. Advanced Chemistry Development, Inc.: ACDLabs logP Predictor, log pK Predictor v. 7.0
8. Perrin D. D., Dempsey B., Serjeant E. P.: „pK<sub>a</sub> Prediction for Organic Acids and Bases”, Chapman and Hall, London 1981.
9. Hansch C., Leo A., Hoekman D.: Exploring QSAR – Hydrophobic, Electronic, and Steric Constants, ACS Professional References Book, ACS, Washington 1995.
10. HyperChem for Windows Release 5.1 Profesional Version, Hypercube Inc.
11. <http://www.logp.com/>.

Received: 13.12.2006